

APPLICATION FOR
UNITED STATES LETTERS PATENT

SPECIFICATION

TO ALL WHOM IT MAY CONCERN:

Be it known that I, **Robert W. Allington**, a citizen of the United States of America, and resident of the State of Nebraska, having a postal address of 2030 Euclid Avenue, City of Lincoln, State of Nebraska, have invented a new and useful "**IMPROVED METHOD AND APPARATUS TO ENHANCE THE SIGNAL TO NOISE RATIO IN CHROMATOGRAPHY**", of which the following forms the specification.

IMPROVED METHOD AND APPARATUS TO ENHANCE THE SIGNAL TO NOISE RATIO IN CHROMATOGRAPHY

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The present application is a continuation-in-part of U.S. Patent Application
Serial Number 10/728,182 filed December 4, 2003, which is a continuation-in-part
10 of S.N.10/636,153 filed August 7, 2003, which is a continuation-in-part of S.N.
10/410,373 filed April 9, 2003, and which are herein incorporated by reference.

BACKGROUND OF THE INVENTION

Field of the Invention

The present invention relates generally to a method and apparatus for increasing
15 the signal to noise ratio of a signal received from a photocell used in capillary High
Performance Liquid Chromatography (HPLC), Capillary Electrophoresis (CE) or
Capillary Electroendosmosis Chromatography (CEC). For instance, in capillary
HPLC a capillary tube serves as the chromatographic column. If the capillary is an
open tube, its inside diameter may be from 10 μ m (detector sensitivity) to 100 μ m
20 (detector volume). Packed capillary columns have analogous volume limits.

More particularly the present invention makes use of a plurality of photocells. In
one embodiment, the photocells are used to detect a solvent spike (absorbance spike
or refractive index spike) of a quiescent fluid. In another embodiment, a set of
photocells receive light through the same particle of fluid by accurately tracking the
25 solvent spike as it passes the linear array of photocells. In either case, the signals
created by all the photocells are summed or integrated over time to increase the signal
to noise ratio. This invention applies to HPLC, CE, and CEC. References herein to
capillary chromatography or capillary HPLC may also apply to CEC and CE.
"Solvent spike" in the claims and elsewhere includes both relating to an absorbance
30 spike as well as refractive index spikes.

Because the present invention is applicable to HPLC, CE, and CEC, these three

will be referred to collectively hereinafter as “capillary separation schemes.” The term “separation schemes” will encompass all the above, plus those used with small volume columns.

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Background Art

To identify components and their concentration in a mixture of solute and solvent using high performance liquid chromatography, light is passed through the solution. The light that is neither reflected nor absorbed impinges on a photocell, where the intensity of the light is converted to an electrical signal. The intensity of the light hitting the photocell is related to the concentration of a particular solute in the solution. Sensitivity of this system is proportional to the path length of the light as it passes through the sample. Recall that the solution is contained in a capillary tube. Increasing the optical path length in the “usual fashion,” however, invites problems related to widening and “blurring” of peaks because of Poiseuille flow distribution or worse, separation, recirculation, or a flow with a helical path, along the light path.

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Sources of noise in the signal include the light source, thermal effects, and turbulence in the flow of the solution. The signal to noise ratio of the signal produced by a single, stationary photocell may be too low to be useful. It becomes difficult or impossible to pick out peaks in the signal because the signal is extraordinarily low and the noise level is as high as in a larger diameter (e.g. 4.7 mm i.d.) chromatographic flow system. A signal to noise ratio of about 2 is considered the lowest acceptable. To overcome this difficulty, it is possible to move the light source and photocell along a capillary tube through which the solute is flowing; or to move the capillary tube past the light source and sensor. The relative velocity at which the capillary tube moves compared to the light source and photocell is equal to the maximum velocity (in any infinitesimally thin slice of fluid in cross-section) of the fluid. The purpose is to generate a “signature” of a particular particle of fluid moving at the centerline speed of the flow. This approach has its weaknesses, including the need for accurately correlating the relative positions of the detector and capillary tube with the peaks observed. The need for moving parts or specially triggered and

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secondarily detected flash from a flashlamp increases the complexity of the apparatus and the potential for failure.

R. E. McKean, in his University of Massachusetts Ph.D. dissertation “Improving the Signal-to-Noise Ratio by Cross Correlation in Flow Injection Analysis and High Performance Liquid Chromatography” (1990) revealed a method for reducing the signal to noise ratio in high performance liquid chromatography. His approach was to produce a clean (relatively noise free) signal in an artificial setting with high concentrations of the solutes. This clean signal was then cross correlated with the noisy signals produced in the usual settings. Although this approach was successful, it brings up the question of how to produce a clean signal when the solutes are unknown. Also, McKean used chromatography equipment of the late 1980’s.

McKean discusses the ensemble averaging of multiple signals to improve the signal to noise ratio. He does not, however, suggest the use of multiple sensors and indicates the averaging approach would be “time consuming” for high performance liquid chromatography, presumably due to needing to run multiple identical samples past a single sensor. McKean, because his research focused on his cross correlation method, was not motivated to utilize multiple sensors for summing or averaging signals in a complete chromatogram.

Another novel approach was suggested by Hjerten in U.S. 5,114,551 in which a single detector was used to pick up a signal at multiple locations on the capillary. This was done by looping the capillary around and returning back to the light source and sensor location. A relatively small improvement in the signal to noise ratio would be realized with this method due to the limited number of chromatograms that can feasibly be taken. In one embodiment of this invention, the capillary tube is moved laterally in order to move a new portion of the capillary tube between the light source and sensor. This is an unnecessary complication, requiring control circuitry and moving parts that can fail. The flow, too, is not favorably enhanced by looping or by moving the capillary tube.

There is, therefore, a need for a way to significantly improve the signal to noise ratio of the signal produced by photocells in capillary high-pressure liquid

chromatography with no moving parts.

Summary of the Invention

A purpose of this invention is to provide a method and device capable of
5 producing a substantially clean signal in high-pressure liquid chromatography.

Another purpose of the present invention is to carry out the aforementioned purpose
with no moving parts. As part of these purposes, an objective of the present invention
is to accurately track a leading edge indicator, such as a solvent spike or opaque fluid
plug as it flows along its capillary tube.

10 It is well known that the sensitivity of a chromatograph, produced by any of the
capillary separation schemes, HPLC, CE, or, CEC, is directly proportional to the path
length of a beam of light passing through the sample. An indirect method of
increasing this path length is to repeatedly take a chromatograph of the same particle
of solution. A linear array of monochromatic light sources and sensor photocells are
15 aligned parallel with a polished quartz capillary tube. Some of the first photocells
encountered by the solvent spike are used to accurately locate the leading edge
indicator, positively identifying a fluid particle. The leading edge indicator may be a
solvent spike or a plug of fluid having significantly different optic characteristics than
the solution and is a relatively large fluid particle about the same size as the injection
20 volume. A solvent spike arises when the injecting solvent has a refractive index
differing from that of the eluting solvent at the same time and location as the
injection. Because the scanning tube is a cylinder, the light seen by each photocell
varies sharply as the solvent spike (refractive index spike) passes them by. It differs
from the absorbance (chromatographic) signal in that the solvent spike is usually
25 taller and arrives first. Therefore it is easy to detect as it is located or goes past any
photocell location. In applications involving CE or CEC, and sometimes also
capillary HPLC, the solvent spike may be replaced by an unretained, or otherwise
leading UV absorbing reference peak. This is accomplished by including a selected
absorbance reference material added to the sample. In a first embodiment of the
30 invention, the flow of the solution is stopped within the capillary tube at a moment

when the leading edge indicator is oriented at a known photocell. The cessation of the flow is accomplished using a three-way valve. At the moment the leading edge indicator is located at a predetermined photocell, such as the 1000th photocell, the three-way valve is actuated to divert the solution to a waste port, thereby relieving the solution in the capillary tube of its pressure. The solution will continue to flow in the capillary tube until the pressure is relieved. Once the solution in the capillary tube is quiescent, the photocell sensors in the linear array are used to scan the solution to obtain a chromatogram for the particular sample. Each photocell will either scan repeatedly, and the scans summed, averaged, or statistically correlated, or the scan will be taken over time and integrated.

In a second embodiment of the invention, the solution flows as usual and a solvent spike in the solution is accurately tracked as it is scanned by a series of photocells in the linear array. Signals from each of the photocells, as the same particle of fluid passes through the associated light beam, are summed or statistically correlated. Because the same particle of solution is being tested, the pertinent information in the signal is the same for each reading. The noise should not be correlated to this method of taking multiple readings. The resulting sum (or average, or statistical correlation) has an improved signal to noise ratio because the signal is strengthened by a factor of N (where N is the number of photocell sensors used to scan a particular fluid particle), while the noise is only strengthened by a factor of \sqrt{N} (assuming white noise).

The novel features believed to be characteristic of this invention, both as to its organization and method of operation together with its further objectives and advantages, will be better understood from the following description considered in connection with the accompanying drawings in which a presently preferred embodiment of the invention is illustrated by way of example. It is to be expressly understood however, that the drawings are for the purpose of illustration and description only and not intended as a definition of the limits of the invention.

Brief Description of the Drawings

Fig. 1 shows a light source, capillary tube and photocell sensors.

Fig. 2 shows a flow chart showing steps for carrying out the present invention with a quiescent solution and taking a continuous reading.

5 **Fig. 3** shows a flow chart showing steps for carrying out the present invention with a quiescent solution and taking multiple readings.

Fig. 4 shows how information is carried from an array of photocell sensors, ultimately to a chromatogram when the solution is quiescent during scanning.

Fig. 5 shows a representative clean signal produced by a linear array of 1024 photocell sensors.

10 **Fig. 6** shows a representative noisy signal produced by a linear array of 1024 photocell sensors.

Fig. 7 shows a cleaned signal produced using the methods of the present invention with a linear array of 1024 photocell sensors.

15 **Fig. 8** is a flow chart showing steps for carrying out the present invention with a flowing solution.

Fig. 9 shows how information is carried from an array of photocell sensors, ultimately to a chromatogram when the solution is flowing during scanning.

Fig. 10 shows a representative clean signal produced over time by a photocell sensor.

Fig. 11 shows a representative noisy signal produced over time by a photocell sensor.

20 **Fig. 12** shows a representative cleaned signal produced over time by a photocell sensor.

Best Mode for Carrying Out the Invention

25 In **Fig. 1** a schematic of an apparatus for the present invention is depicted. A uniform, monochromatic light source (or sources) **100** lines one side of a polished, quartz capillary tube **110**. Directly opposite (on the other side of the capillary tube **110**) is a linear array of photocell sensors **120**. (Only twelve individual photocell sensors **141–152** are shown in **Fig. 1**, however, in practice many more individual
30 photocell sensors **141–152** would be used.) Some of the light emitted from the light

source **100** is reflected off the capillary tube **110** and the solution flowing through the capillary tube. Some of the light is absorbed by the solution. That light not reflected or absorbed, passes through the capillary tube **110** and the solution flowing in the capillary tube. Each of the photocell sensors **141–152** creates a signal related to the light intensity of the light that impinges on it. An identifying feature of the components of the solution is the amount of light absorbed.

A signal of interest is one over a period of time. A fluid particle **130** is defined as a small mass of fluid of fixed identity. As a fluid particle **130** of the solution is scanned by a photocell **141–152**, a signal is recorded based on the light passing through the fluid particle **130** and impinging on the photocell sensor **141–152**.

In the first embodiment of the present invention, the fluid is quiescent when the scanning step is carried out. Therefore, any given photocell **141–152** records data for a single fluid particle **130**. The data recorded might be an integral of each individual photocell's **141–152** continuous signal over time, or a summation, average or statistical correlation of a series of the photocell's **141–152** signals taken sequentially over time. If the readings are taken sequentially, the frequency at which readings are taken must be greater than the frequency of any significant high frequency noise. The total time over which readings are taken must be several times greater than any significant low frequency noise. This total time will be on the order of 10-20 seconds, and preferably no longer than a minute in most cases.

After readings are taken from each photocell **141-152**, the integrated, averaged, or correlated signals are concatenated into a new signal having a greater signal to noise ratio than the original.

To halt the flow, a three-way valve **160** is provided at an inlet to the capillary tube **110**. An indication **155** of the leading edge of the chromatograph sample, such as a solvent spike or a fluid plug having significantly different optic characteristics than the solution of interest (an opaque fluid plug is an example), is tracked by its signature on the photocells **141-152**. When the leading edge indicator **155** reaches a predetermined photocell **141-152**, such as the 1000th photocell, the solution entering the three-way valve **160** at its inlet **170** is redirected by the three-way valve to the

inlet waste port **180**. Because the fluid has a finite bulk modulus of elasticity, the fluid may continue to flow before it stops as the pressure is relieved in the capillary tube **110** so that the leading edge indicator may travel beyond the 1000th photocell. Some of the sample may exit the capillary tube **110** at the outlet waste port **190**. When the pressure is relieved, the flow will cease.

In the second embodiment of the present invention, the same particle of fluid **130** travels past each of the photocell sensors **141–152** in turn. The velocity of the fluid particle **130** is determined as follows. A location of a solvent spike is detected using the first photocell sensors **141–152** the solvent spike encounters. For instance, 1000 photocell sensors **141–152** in an array **120** of 2048 photocell sensors may be used to detect a solvent spike. It is known that the solvent spike is a fluid particle about the same size as the injection volume. The solvent spike is formed when the injecting solvent has a refractive index differing from that of the eluting solvent at the same time and location as the injection. The light detected by each photocell sensor **141–152** varies sharply as the solvent spike (refractive index spike) passes between the light source **100** and a photocell sensor **141–152**. The signal produced by a photocell sensor **141–152** when the solvent spike is scanned differs from the absorbance (chromatographic) signal in that the signal due to the solvent spike is usually taller and arrives before other peaks. The said spike may also be an absorbance spike. Once the spike has been detected, its velocity is determined using a small number of additional photocell sensors **141–152**, for instance, as the spike passes from the 1000th photocell sensor to the 1005th photocell sensor. This velocity, in photocell sensors per unit time, is multiplied by a predetermined factor, for example 1001, to obtain a scanning speed, again in photocells per unit time.

Using this scanning speed, a number of photocell sensors **141–152** equal to the predetermined factor (e.g. 1001) are scanned. Using the example, above, the 6th through the 1006th photocell sensors are scanned. At the end of this scan, the solvent spike is again located using the data just obtained from the photocell sensor scan. The solvent spike should be at the last photocell sensor from which data were obtained. According to our example, the solvent spike should be located at the 1006th

photocell sensor. If the solvent spike is not located at the correct sensor, the solvent spike velocity is recalculated from the new data, a new scanning velocity is calculated and the process repeated using the correct bank of photocell sensors **141–152**. In our example, this new correct bank of photocell sensors **141–152** for the next step would be the 7th through the 1007th photocell sensors.

Figs. 2 and 3 depict flow diagrams for the first embodiment wherein the solute is quiescent when scanned. Initially, the solute is pumped into the capillary tube **110**. The location of the solvent spike or other leading edge indicator **155**, such as a plug of fluid having different optic qualities than the solution, is detected **200** as the solute flows in the same manner as described above. When the solvent spike arrives at a predetermined photocell sensor **141–152**, for example the 1000th in an array **120** of 1024 photocell sensors, the solute flow is stopped **210**, by stopping the pump, or by closing a valve, or redirecting the flow to the inlet waste port **180** with a three-way valve **160**. With the solute in a quiescent state, and the solvent spike located at a known photocell sensor **141–152**, all the photocell sensors **141–152** in the array **120**, or a predetermined subset of the photocell array **120**, are scanned continuously **220** (**Fig. 2**) for a predetermined duration sufficient to suppress all the significant low frequency noise. The result of this continuous scan is integrated **230** in the photocell sensor **141–152** if its charge storage capacity is adequate, or in a separate integrator for each photocell sensor **141–152**. The resulting information is a plot of the light passing through the capillary tube **110** with the independent variable (abscissa) being the photocell number from which the signal was taken (see **Fig. 7**). This abscissa can easily be converted to the location along the capillary tube **110**, x , if preferred.

In **Fig. 3**, the same approach as that shown in **Fig. 2** is taken, except that instead of a continuous scan **220**, scans are taken repeatedly **320** and then summed, averaged, or statistically correlated **330** to produce the reduced-noise signal. The frequency at which these sequential readings are taken must be sufficiently high for the suppression of significant high frequency noise. The total time over which the sequential readings are taken must be adequate to suppress significant low frequency noise.

The process of scanning, calculating, and storing the data is depicted in **Fig. 4**. The linear photocell sensor array **120** is shown at the top with twelve photocell sensors **141–152** shown. The ellipses shown to the left of photocell sensor **141** and to the right of photocell sensor **152** indicate there may be more photocell sensors.

5 The signal from each of the photocell sensors is fed into a set of processing blocks **440–453**. These processing blocks **440–453** may include an Analog to Digital (A/D) converter and an integration, summation, or statistical correlation function. The processing blocks **440–453** may be inherent to the photocells, themselves, if the charge storage capacity of each photocell sensor is adequate for the task, or they may
10 be separate units, carrying out their operations in analog or digital mode. Finally, the resulting, processed data are organized into a chromatogram, as indicated by the plot **470** shown.

A representative plot of a scan is shown in **Fig. 5**. This plot shows a noise-free signal of the quiescent solution as taken from 1024 photocells. Here, four peaks or
15 spikes are shown. On the abscissa is the photocell sensor number from 1 to 1024, while the ordinate is the signal, as amplified from the photovoltaic sensors, in volts.

In **Fig. 6** the noise-free signal is shown with white noise superimposed upon it, resulting in a noisy signal. The white noise has a maximum amplitude of three volts. The clean signal cannot be identified due to the noise.

20 In **Fig. 7**, 100 noisy signals like that shown in **Fig. 6** have been integrated with the time of integration divided out, or arithmetically averaged, or statistically correlated. As can easily be seen in the plot **470**, due to the averaging step, the relative fraction of the signal attributable to noise is greatly reduced.

A flow diagram of the second embodiment of the present invention is shown in
25 **Fig. 8**. Once the solvent spike is detected **200**, its speed, V_{ss} , is estimated **800** at photocell sensor m , such as the 1005th photocell sensor, as used in the example, above. Then the n ($n = 1001$ in the above example) photocell sensors from $m+1$ “back” (upstream) are scanned at a rate, V_{sr} , such that, when the scan is finished, the solvent spike should have reached photocell sensor $m+1$ **810**. The scanning speed is
30 calculated as $V_{sr} = nV_{ss}$, where both V_{sr} and V_{ss} are in photocells per unit time.

After the aforementioned scan **810**, the location of the solvent spike is, again, detected **820**, ultimately to ascertain that it did, in fact, reach photocell sensor $m+1$, and no further. Before comparing the location of the solvent spike to photocell sensor $m+1$, the value of m is incremented up by one (1.0) **830** and this new value of m is tested **840** against N , the total number of photocell sensors **141–152** (2048 in the example above), so the process ends when the last photocell sensor is encountered. If $m \leq N$ at this point, the location of the solvent spike is compared **850** with the location of the photocell sensor m . If the solvent spike is at photocell sensor m , the same estimated solvent spike speed is used and the process repeated, scanning n photocell sensors upstream from and including the new photocell sensor $m+1$ **810**. If the solvent spike is not at photocell sensor m , a new solvent spike speed is estimated **800** before the remainder of the process is repeated as before.

For the present embodiment wherein measurements are made of the flowing solution, the photocell sensors **141–152** are again shown in **Fig. 9** with ellipses shown at each end of the linear array **120** to indicate there may be more photocell sensors than shown. The analog signals from the photocell sensors **141–152** are converted to digital signals in an A/D converter **900**. Because the front of the n photocell sensors is shifted such that it moves with the flow, and only n photocell sensors are read at each scan, the digital signals, from the first to the last, need only to be stored in memory locations **941–952** from the first to the last. No more shifting is required.

From the memory locations **941–952**, the signal is processed in a calculation function **910** that integrates, sums or statistically correlates each photocell sensors' **141–152** signal to produce a chromatogram, as indicated by the plot **970**.

A chromatogram, such as would be produced by the second embodiment of the present invention, is shown in **Fig. 10**. **Fig. 10** represents a clean (noiseless) HPLC with four peaks. Although the abscissa could be the photocell numbers of the n photocell sensors used for each scan, it is just as logical to make the abscissa be time in seconds. The ordinate is, again, the signal, as amplified from the photovoltaic sensors, in volts.

The same signal as shown in **Fig. 10** is replotted in **Fig. 11** with simulated noise

superimposed on the clean signal. The white noise has a maximum amplitude of three volts. The clean signal cannot be identified due to the noise.

The next step in the analysis is shown in the plot 970 of Fig. 12. Here, 100 noisy signals, with the same clean content as shown in Fig. 10 and different noise (all with a maximum amplitude of three volts), were averaged. The improvement can easily be seen when comparing Fig. 12 with Fig. 11. The improvement is evident, even though only 100 readings were used (in practice, many more could be used). Even the last and smallest peak (seen in Fig. 10 at about 515 seconds) can be resolved from the noise. The signal could also have been summed or statistically correlated.

Obviously many modifications and variations of the present invention are possible in light of the above teachings. Any number of photocell sensors may be used in the linear arrays. The photocell sensor array need not be linear. It is, therefore, to be understood that within the scope of the appended claims, the invention may be practiced otherwise than as specifically described.